

Yeast homologues of higher eukaryotic TFIID subunits

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ABSTRACT In eukaryotic cells the TATA-binding protein (TBP) associates with other proteins known as TBP-associated factors (TAFs) to form multisubunit transcription factors important for gene expression by all three nuclear RNA polymerases. Computer searching of the complete *Saccharomyces cerevisiae* genome revealed five previously unidentified yeast genes with significant sequence similarity to known human and *Drosophila* RNA polymerase II TAFs. Each of these genes is essential for viability. A sixth essential gene (*FUN81*) has previously been noted to be similar to human TAF_{II}18. Coimmunoprecipitation experiments show that all six proteins are associated with TBP, demonstrating that they are true TAFs. Furthermore, these proteins are present in complexes containing the TAF_{II}130 subunit, indicating that they are components of TFIID. Based on their predicted molecular weights, these genes have been designated *TAF67*, *TAF61* (68), *TAF40*, *TAF23* (25), *TAF19* (*FUN81*), and *TAF17*. Yeast *TAF61* is significantly larger than its higher eukaryotic homologues, and deletion analysis demonstrates that the evolutionarily conserved, histone-like domain is sufficient and necessary to support viability.

The TATA-binding protein (TBP) is a key element in transcription by all three eukaryotic RNA polymerases (1). TBP binds a set of TBP-associated factors (TAFs) to form the transcription factor TFIID, the core factor involved in RNA polymerase II transcription (2). Other subsets of TAFs associate with TBP in different complexes (SL1, TFIIB, SNAPc) involved in production of RNA polymerase I, RNA polymerase III, and snRNA transcripts, respectively (1). In addition, there is evidence that TBP associates with various groups of TAFs to form TBP complexes distinct from TFIID but of undetermined function (3, 4).

Most information about the function of TAFs has been obtained from *in vitro* studies on partially purified mammalian and *Drosophila* factors (5). These studies suggest that TAFs are essential for the response to transcriptional regulatory proteins but not for basal transcription *in vitro* (6). More recently, several TAFs have been identified in yeast extracts (7–9) and studied *in vivo* (10, 11). These *in vivo* analyses have shown that TAFs are important for transcription of specific genes but are not universally required for transcriptional response to activator proteins. In particular, studies both in yeast (11, 12) and in a hamster cell line with conditional expression of TAF250 (13) implicate TAFs in the correct expression of cell-cycle genes. Other experiments show TAFs to be important for the expression of some genes whose transcription is directed by weak TATA elements or initiator elements (10, 14). Taken together, the results of these *in vitro* and *in vivo* studies suggest that TAFs play important gene-specific roles in transcription and are potential, but not obligate, targets of trans-activating proteins.

Yeast TBP was originally purified as a single protein (15–18). This stands in contrast to its mammalian counterpart, which remains tightly associated with TAFs throughout mul-

iple chromatographic purification steps. However, affinity purification experiments revealed the presence of a constellation of factors associated with TBP in yeast (7, 8). Cloning of the genes for several of these proteins revealed them to be highly similar in sequence to the known mammalian and *Drosophila* TAFs. Thus far, genes have been identified that encode yeast TAF130/145, TAF90, TAF60 (7, 8), and TAF25 (9). In addition, an open reading frame with unknown function (*FUN81*; ref. 19) has been noted to resemble human TAF_{II}18 (20). However, this yeast protein has not previously been shown to associate with TBP.

Assuming the composition of the yeast TFIID complex to be similar to that of mammalian and *Drosophila* TFIID, we expected that there would be yeast homologues of the other known higher eukaryotic TAFs. The completed sequencing of the *Saccharomyces cerevisiae* genome (21) allowed us to search for potential homologues of the remaining known TAFs by protein sequence similarity. Here we report the identification of several previously unknown yeast proteins as true yeast TAFs associated with the TFIID complex.

MATERIALS AND METHODS

Sequence Analysis. Protein sequence similarity searching was performed by BLAST (22) search of the complete yeast genome [*Saccharomyces* Genome Database (SGD), Stanford University]. Alignments were made using the GCG and SEQVU programs.

TAF Gene Cloning. The putative TAF genes were amplified by PCR using Vent polymerase (New England Biolabs) and cloned into yeast shuttle vectors using standard molecular biology techniques (23).

Gene Disruptions. Candidate TAF gene deletions were performed in the diploid yeast strains YSB286 (*MATa/MAT α* , *ura3-52/ura3-52*, *leu2 Δ 1/leu2-3*, 112, *his3 Δ 200/his3 Δ 200*, *TRP1/trp1 Δ 63*) or KY320 (*MATa/MAT α* , *ade2-101/ade2-101*, *his3 Δ 200/his3 Δ 200*, *leu2::PET56/leu2::PET56*, *trp1 Δ 1/trp1 Δ 1*, *ura3-52/ura3-52*) by standard one-step gene disruption-deletion using either *LEU2* or *ADE2* as the disrupting marker (24). Disruption of one copy of the gene of interest was verified by Southern blotting in all cases. The heterozygous diploids were sporulated and tetrads were dissected onto rich growth medium to determine viability of the resulting haploids. Details of the disruption constructs will be provided on request.

Creation of Epitope-Tagged Alleles. The plasmid ZM253 (*HIS3*, *CEN/ARS*), containing the inducible *GAL1* promoter upstream of three copies of the influenza hemagglutinin (HA) epitope (25), was constructed to allow immunological detection of the TAF proteins. Each putative TAF protein coding sequence was cloned into this vector in frame with the triple epitope tag. The resulting plasmids were each introduced into a haploid strain containing a chromosomal deletion of the same gene using plasmid shuffling (26) on plates containing 2% galactose and 5-fluoro-oroic acid (FOA). In each strain, the chromosomal deletion of the gene was fully complemented

by the epitope-tagged copy of the gene. All strains required galactose for growth.

Immunoprecipitations and Western Blotting. Each strain expressing an epitope-tagged TAF homologue (or epitope-tagged Fus3 as a control) was grown to mid-logarithmic phase in 100 ml of YP medium (1% yeast extract/2% bacto-peptone) containing 2% galactose and 0.2–0.5% dextrose. Cells were harvested and whole-cell extracts were made by glass bead lysis in 450 mM Tris-acetate (pH 7.8), 150 mM potassium acetate, 60% (vol/vol) glycerol, 3 mM EDTA, 3 mM dithiothreitol, and 1 mM phenylmethylsulfonyl fluoride (27). In early experiments, yeast whole-cell extracts were further processed by chromatography on BioRex 70 (Bio-Rad) as described (27), and the BioRex 70 0.6 M eluate was used for immunoprecipitations (see Fig. 3, TAF17 and TAF40). This fractionation was subsequently found to be unnecessary, and all further experiments were performed with unfractionated whole-cell extract. TAF17 and TAF40 immunoprecipitations from whole-cell extract yielded the same result as shown in Fig. 3.

Immunoprecipitations were performed according to a modification of a protocol provided by N. Kuldel (personal communication). Reactions contained 100–300 μ g of protein extract. The sample volumes were adjusted to 500 μ l with buffer A [20 mM Hepes, pH 7.6/20% (vol/vol) glycerol/1 mM dithiothreitol/1 mM EDTA] containing 125 mM potassium acetate, and Nonidet P-40 was added to a final concentration of 1%. The protein extract was precleared by incubation for 1 hr with 50 μ l of preswollen 10% (vol/vol) Protein A Sepharose beads (Sigma) in buffer A. Protein A beads were coupled to either rabbit polyclonal α -TBP or to rabbit preimmune serum as described (28). Reactions of extract with 50 μ l (10%, vol/vol) of coupled beads were allowed to proceed overnight at 4°C on a rotator apparatus. Antigen–antibody complexes were recovered by centrifugation and were washed five times with 1 ml of buffer A containing 125 mM KOAc and 1% Nonidet P-40. Samples were boiled in SDS/PAGE loading buffer and subjected to SDS/PAGE analysis.

Proteins were resolved on 10% or 12% acrylamide gels and then electroblotted to nitrocellulose. Antibody probing and washing was performed according to standard techniques (28), and enhanced chemiluminescent detection was used to visualize bands as recommended by the manufacturers (Amersham; Kirkegaard & Perry Laboratories).

Deletion Analysis of TAF61. Deletion alleles of *TAF61* were constructed using both PCR cloning and existing restriction sites in the *TAF61* gene and were inserted into a yeast *TRP1*, *CEN/ARS* plasmid. The plasmids were transformed into a haploid strain containing a chromosomal disruption of *TAF61* complemented by a *URA3* plasmid bearing the wild-type *TAF61* gene. The resulting transformants were transferred to FOA-containing medium (26) to determine whether each deletion plasmid could support viability.

RESULTS

Yeast Homologues of Human and *Drosophila* TAFs. Protein similarity searches (22) of the complete *S. cerevisiae* genome were carried out using all published mammalian and *Drosophila* RNA polymerase II TAFs as query sequences. Each search revealed a unique yeast sequence with significant similarity to the known mammalian or *Drosophila* TAF. The one exception was *Drosophila* TAF_{II}110 (29, 30), for which no similar yeast sequence was found. For simplicity, we have designated each gene according to the predicted molecular weight of the encoded protein. Sequence alignments are shown in Fig. 1.

Human TAF_{II}55 (31, 32), for which no *Drosophila* homologue has yet been described, is similar to a yeast open reading frame encoding a protein of predicted molecular mass 67 kDa (Fig. 1A). Relative to the human protein, the yeast protein (SGD accession no. YMR227C) has additional nonhomolo-

gous protein sequences at both the amino and carboxyl termini. Interestingly, the amino-terminal extension is rather basic, while the carboxyl-terminal extension is quite acidic.

Searches with human TAF_{II}28 (20), which has been implicated in the transcriptional response to retinoic acid receptor (33), revealed a predicted yeast protein of 40 kDa (SGD accession no. YML015C). The same yeast protein was also matched with searches using the *Drosophila* homologue TAF_{II}30 β (34). The yeast protein is slightly larger than its counterparts, with the nonoverlapping sequences appearing at the carboxyl terminus of the protein (Fig. 1C).

Human TAF_{II}30 occurs in a subset of TFIID and is thought to be a potential target for the estrogen receptor AF-2 activation domain (35). No *Drosophila* homologue is known, but a sequence search against the yeast genome uncovered a predicted protein of 23 kDa (SGD accession no. YDR167W) with significant similarity to the human protein. This protein was recently independently shown to be a component of yeast TFIID (9) with an observed molecular mass of 25 kDa and was thereby designated TAF25. The alignment of the human and yeast proteins is presented in that paper.

Several of the higher eukaryotic TAFs show some sequence similarity to histone proteins (2). The relevance of these alignments is supported by structural studies showing that the histone-like regions of *Drosophila* TAF62(60) and TAF42(40) adopt a histone-fold arrangement resembling the histone H3/H4 pair (2). Although a yeast homologue of *Drosophila* TAF62(60) has been described (7, 8), none have previously been reported for the other histone-like TAFs. Similarity searches with dTAF_{II}42(40) or its human homologue TAF_{II}32 match a yeast open reading frame with a predicted molecular mass of 17 kDa (SGD accession no. YMR236W). Interestingly, the sequence similarity includes the histone H3-like domain and an additional region of strong identity between species (Fig. 1D). However, the yeast protein completely lacks a carboxyl-terminal region that is not conserved between the human and *Drosophila* proteins.

The putative histone H2B-like TAFs (human TAF_{II}20 (20, 36) and *Drosophila* TAF_{II}30 α or 22/28 (34)) match a predicted yeast protein of 61 kDa (SGD accession no. YDR145W). We note that this protein has been independently cloned as a component of TFIID with an observed molecular mass of 68 kDa (11). The yeast protein is significantly larger than its higher eukaryotic homologues, primarily due to an extra \approx 300 aa in the amino-terminal half of the protein. This extra domain is predicted to be very hydrophilic and contains several polyglutamine stretches (Fig. 1B).

Candidate Yeast TAFs Are Essential for Viability. To determine whether the new genes were essential for cell viability, we performed one-step gene disruptions in diploid yeast. Dissected tetrads from all strains yielded two live and two dead spores each (Fig. 2 and data not shown), with the surviving haploids always lacking the disruption marker. Therefore, each of the five yeast TAF homologue genes is essential for viability.

Candidate Yeast TAFs Are Associated with TBP. Immunoprecipitations with each of the putative yeast TAFs were performed to determine whether they were indeed associated with TBP *in vivo*. A HA epitope-tagged allele of each gene was created and introduced into a haploid yeast strain as the sole source of the given protein. Immunoprecipitations were performed by incubating yeast extracts with anti-TBP antibody-coupled beads or with preimmune serum-coupled beads. Bound proteins were subjected to Western blotting using a monoclonal antibody (12CA5) directed against the HA epitope (25). As shown in Fig. 3, each HA-tagged protein was found to be specifically associated with TBP, indicating that these proteins are all truly TAFs. Also tested in this experiment was the essential yeast gene *FUN81* (19), which was previously noted to be similar to human TAF_{II}18 (20), but which had not been shown to be associated with TBP. In light of its efficient

A.

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yTAF67      1  MAVIRIKKPRGGPEKEDQPLEGEPKLBIBIKTK
yTAF67     34  VTDEDIKPKPKLKLINLKKKESADGKEKKNLSLK
yTAF67     67  LKLNLKKNNEEPVKIKIHKAPKLBKPIBIPGEAY
yTAF67    100  DSEADIDDDPLIESGVLLPDIQLEFYKNS
hTAF55      1  MSKSKDAPHEESQFPEEY-ASTVARRA
yTAF67    133  LESGDYS--GSIKWK-KNERFAVYTDVYMYG
hTAF55     32  VQSGHVNLDKDRLELHPDGGVRYRVPPLA
yTAF67    162  IIVDITVIVNVNRSYVHKNLITFDVSGMLC
hTAF55     65  SKLVYDLCVMESLKTIDKKTFTYKTDAGQMLVS
yTAF67    195  IRPIQEEEEVYALEAPDTEDLVVKHFEGETE
hTAF55     98  -----TVDGDD
yTAF67    228  WENKRTFLKGYNGAPLSDMEKHLKEIALGYD
hTAF55    104  YPPVVE-----PVASTDPKAKKKDKDKEKFI
yTAF67    261  YKGGGKPLKYNVNRFRFRKMDPNEIDY-VE
hTAF55    132  WNGGKPLKKNVNRFRFRKMDPNEIDY-VE
yTAF67    292  VDMKQKQKQEEVSYDLDKSL-ORQER-
hTAF55    165  EMKRLSTAEAVRTRWEIAEDTKAEENQGL
yTAF67    323  -SSWENFKEEPPEPLSRPAKKEEHTASAV
hTAF55    198  DSS-----PGMSHRQGHDSLHEHRELFNDL
yTAF67    355  GKQGAKEGEGMEFEEDLGAAFSEEEG
hTAF55    227  SSSSEKRETOHOHEEINIITEEDLRQLQD
yTAF67    388  SGAGGKKEGQEVGDEVDQDTGGEDDDDD
hTAF55    260  KLNLSLEQHOHEEENGLVMIQNOIDN----
yTAF67    421  DIEAAGGESDDEKDNRQHTLELADENLE
hTAF55    288  -----MKGKLCETQ
yTAF67    454  TTLAHTKHKSKATPLKSRFIDSKKKEKA
hTAF55    297  DRAKRQEDLMMVEMLAKNRQAVLDEKQKE
yTAF67    487  LKRRKQQTEDSVKQKHGRSDAETANNVEE
hTAF55    330  BREKEQSSLOEELSLLEKE
yTAF67    520  EEEEEEEEEDEVDDEEDDEENDEONVHER
yTAF67    553  EHIQENKVVRELDEAPEETLDQNDLMMMLFG
yTAF67    596  AEGDE
    
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B.

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yTAF61      1  MSSNPENSGVNNANNNTGTGNADAITGAQQNMVL
yTAF61     34  QPRQLQEMAAKFRLLTEARNVGETTPRGKELM
yTAF61     67  FQAAKIKQVYDALTLNRRRQQAQAYNNTSNSN
yTAF61    100  SSNPASIPTEENVPNSSQQQQQQQQQTRNNSNKF
yTAF61    133  SNMIKQVLTPEENQEYKELWQNFQVRHTSIKEK
yTAF61    166  ETYLKQNI DRLEQEI NKQTDEGPKQQLQEKKIE
yTAF61    199  LLNDWKVLKIEYTKLFNNYQNSKKTFFYVECARH
yTAF61    232  NPALHKLFLQESTQQQRVQQQRVQQQQQQQQQQ
yTAF61    265  QQQQQQQQQQQQRQGNQRKISSSNSTEIPSVT
yTAF61    298  GPDALKSQQQQNTIATNPRGNVNTSATEQS
hTAF20      1  MSDLFTDFDSNGVARHHLHNNHNS
dTAF30 α    1
yTAF61    331  KAKVINVNATASMLNINISSKSAIFKQTEPAIP
hTAF20      1  MNQFGP-----AL I
dTAF30 α    25  TSSASGLLHDPPIASPSQH-----PMT
yTAF61    364  ISENIISTKTAPVAYRSNRPTTGGSAMNSAL
hTAF20     11  NLSNFSI KPEAPASTPPQ-IGSMANSTAVVKIPG
dTAF30 α    48  NNSNSSSQNGGPFYGLGT-GGPISSGGSKSNH
yTAF61    397  NTPATTKLPPFYEMDQORVMSRKLRELVTGVI
hTAF20     43  TPGAGGRLSL-----ENNVLTMLKLDLV
dTAF30 α    80  TSSAAGS-----ENTPMLTKPRTELV
yTAF61    430  DRDGETVLDGDEVELLDLADDFVTNITAFSC
hTAF20     69  EVDPNQOLDEDEVEMLQADDFLESIVAAAG
dTAF30 α   103  EVDITTOLEDEVEELLCITDQDFVRLTKSIS
yTAF61    463  RLAKHRKSNLEARDLQCHERNWNHFRPQVSA
hTAF20    101  QLAFHRKSNLEARDLQCHERNWNHFRPQVSA
dTAF30 α   135  AFAKHRKSNKLEARDLQCHERNWNHFRPQVSA
yTAF61    496  DETRSTRKRNWPSQNYNCKLQSLT-SDPVA AAK
hTAF20    134  DELRYPYKACTTEAHKQRMALRKTIK
dTAF30 α   168  DELRYPYKAAVTEAHKQRMALRKTIK
yTAF61    527  NNGNNVASLNTKK
    
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C.

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yTAF40      1  MTEPGLDTPKVNYPPLTIANFYSTK
hTAF28      1  MDDAHESSDKGGGETGESDETAAPVPGD--GAT
dTAF30 β    1  MDIEIFPPTQKSNSSDGDVDVDFKFFQS--ASG
yTAF40     30  QMIQVIEEDDDVYVWKQNLHGGTSINQLN
hTAF28     32  D-TDGPPEITDGDADVDLKEAAEEGELESG--
dTAF30 β    32  ERKISDTSIPGNDADRDQKQADGDNDKNTD--
yTAF40     63  KYPKYKYQKTRINQQPDSNKNVPELIFRQD
hTAF28     62  -----DVSQLTVEREDSSLNFAAK
dTAF30 β    63  -----GDGDSG-----EFAHK
yTAF40     95  IQQQIQNSNYEDTNTNEDENKLAQDEQFKL
hTAF28     83  KKKIDKKEKKEKQKVDEDEIKM-----QLL
dTAF30 β    74  KKK-----TKKELLEEFERFM-----QVL
yTAF40    128  VTNLDKQDITNREVEVHRTSLNRQVYKASVVA
hTAF28    110  VSSIFSEELNRYEMRRSAFPKAAUKLIIQSTT
dTAF30 β    93  VSNFTIEEQLDRYEMRRSAFPKAAVKRMOIT
yTAF40    161  NQSSSENRVYFQAAGRLVAGELALAMIKNK
hTAF28    143  GTSVSOKVVAAMSGSKVEVGEVVEEALDY--
dTAF30 β   126  GCSVSONVVAAMSGAKVYVGEVVEEALDY--
yTAF40    194  WLTSQMCIEFDRRTKIQYKLRKYLKLTFSIE
hTAF28    173  -----CEKXWGEIMPPLQPKHREAV----
dTAF30 β   156  -----MGAQGESALQPKHREAV----
yTAF40    227  NQQYKQDYKSDSVPEDHDFRYFDDEEVDKRETT
hTAF28    192  -RRLKSKGGIPNSKHKKLIIE
dTAF30 β   175  -RRLKTKDRMPIGRYQGVRLN
yTAF40    260  LGNSLLQSKSLQSDHNSQDLKQLIEQYNKLV
yTAF40    293  LQFNKLDVSI EKYNNSPLLPEHIREAWRLYRLQ
yTAF40    326  SDTLPNAYWRQTGGEGGSGMFR T
    
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D.

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yTAF17      1  MNGGGKVLNKNVGSVSEVGPDSSTQEEETRHOV
hTAF32      1  MESIGKT--ASPKSMPKDA
dTAF40      1  MSAEKSIDAKAKISAQIKHVKKDA
yTAF17     34  RLCHLIRASQSIHOVEIDPPLQDMDFAHFYIQG
hTAF32     17  QMMAQLKQDMGUTEPYPRMINDMEEPAFRYVIT
dTAF40     23  QVIMSLKELNVDVEYPRVINQLLEFRFRVITC
yTAF17     67  VRKQALVYNDYAGSGNSAGSGEGVGDITRAAA
hTAF32     50  LDDAKLIVSSSHAKK-----ATVDADDVRLAQ
dTAF40     56  LDDAKLVANHAHK-----KTDLDQVRLATEV
yTAF17    100  RTQYQKPTAPKELMQLAARNKKAALPOVMGT
hTAF32     78  RADQSSTSPLEPDLIARQNRQTPPLTKPKPY
dTAF40     84  TLDKISFQJLEHVAKVAADVNSMPLPKPKPH
yTAF17    133  WGVRLPPEKYCLAKKWDL--GDKSKM
hTAF32    111  SGPRLPDPDRYCLAPRYRLKSLG--JKKASTSAG
dTAF40    117  CGLRPLPDRYCLGVRYKLRATNPKPKMTKSAV
hTAF32    142  RITVPRLSVVG-----SVTSRPSTPTLGTPTP
dTAF40    150  EGRPLKTVKPVSSANGPKRPHSVVAKQVVTI
hTAF32    168  QTMSVSTKVGTPMSLTDQRFTVQMPSTSPAVK
dTAF40    183  PKPVIKFTTTTTKTKVQSSGGSGGGGGGEVKSE
hTAF32    201  ASIPATSAVQNVLINPSLIGSKNILITNMMSS
dTAF40    216  STGAGGDLKMEVDSDAAVVGSAGASGSA
hTAF32    234  QNTANESSNALKRRKREDDDDDDDDDDYDNL
dTAF40    249  SGGGGGGGSSGVGVAVKREEREFEFFVTI
    
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FIG. 1. Candidate yeast TAF proteins are similar to known human and *Drosophila* TAF proteins. (A) Yeast TAF67. Basic residues in the amino-terminal region are underscored; acidic stretches in the carboxyl-terminal region are underlined. (B) Yeast TAF61. Polyglutamine stretches are underscored. (C) Yeast TAF40. (D) Yeast TAF17.

interaction with TBP, we now refer to this protein as TAF19, in accord with its predicted molecular weight. As a negative control, extract from a strain containing an epitope-tagged Fus3 protein (37) was immunoprecipitated in parallel and failed to bind TBP.

We also performed the converse experiment, in which 12CA5-coupled beads were used for the original immunopre-

cipitation and then Western blots of the bound proteins were probed with anti-TBP antibody. Again, this experiment shows each of the new proteins to be associated with TBP (data not shown).

Under our experimental conditions, TAF67 appears weakly associated with TBP and requires significantly longer exposures to be detected. Reprobing of the Western blot in Fig. 3

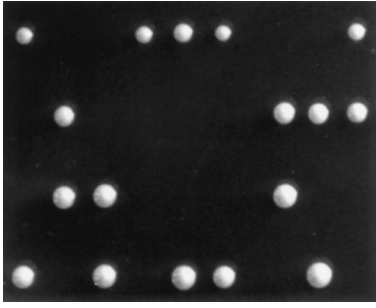


FIG. 2. Candidate TAF genes are essential for yeast viability. The four dissected spores from each tetrad of a *taf61Δ256::LEU2/TAF61* diploid yeast strain are arranged vertically. Identical results were obtained with deletions of all other candidate TAFs (data not shown).

with anti-TAF130 antibody indicates that TAF130 is present in equal amounts in all immunoprecipitations (data not shown), so that the lower affinity of TAF67 for TBP is not a result of failure of this particular immunoprecipitation. We cannot distinguish from this experiment whether there is actually a lower relative amount of TAF67 associated with TBP *in vivo*, or whether the TAF67 interaction is particularly weak and susceptible to disruption under our experimental conditions.

Candidate Yeast TAFs Are Components of TFIID. Having determined that the new genes encode true TAFs, we set out to determine whether these TAFs might, like their mammalian and *Drosophila* counterparts, be part of the TFIID complex. To this end, we probed 12CA5-precipitated proteins from each HA-tagged TAF strain with antibodies directed against TAF130, which has been shown to be an important scaffold for the assembly of TFIID. As shown in Fig. 4, this large TAF is associated with each epitope-tagged TAF, suggesting that the smaller TAFs are part of the TAF130-nucleated TFIID complex.

The Conserved Region of Yeast TAF61 Is Sufficient and Necessary to Support Viability. As mentioned above, yeast TAF61 is significantly larger than its human or *Drosophila* homologues, with the homology being entirely confined to the carboxyl terminus of the yeast protein. This domain includes the region similar in sequence to histone H2B. To determine

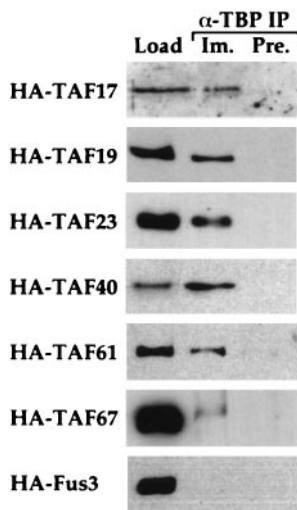


FIG. 3. Candidate yeast TAFs are associated with TBP. Extracts were prepared from strains carrying epitope-tagged alleles of the indicated proteins and immunoprecipitated with anti-TBP antibodies. The TBP-associated proteins were blotted and probed with the monoclonal antibody recognizing the epitope tag. Lanes: Load, 30 μ g of total extract; Im., immunoprecipitate from 300 μ g of total extract with anti-TBP; Pre., immunoprecipitate from 300 μ g of total extract with preimmune serum. Fus3 serves as a non-TAF negative control.

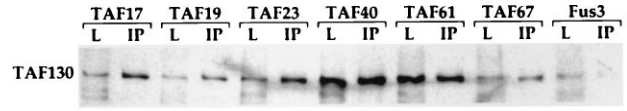


FIG. 4. Candidate yeast TAFs are associated with TFIID. Extracts were prepared from strains carrying epitope-tagged alleles of the indicated proteins and immunoprecipitated with the 12CA5 monoclonal antibody. To determine whether the candidate yeast TAFs were associated with TFIID, the precipitated proteins were blotted and probed with anti-TAF130. Lanes: L, 15 μ g of the indicated extract; IP, immunoprecipitate from 150 μ g of extract. Fus3 serves as a non-TAF negative control.

the importance of the nonhomologous region, we created several truncated alleles of this TAF and used plasmid shuffling to analyze their ability to promote normal cell growth. As shown in Fig. 5, the amino-terminal two-thirds of this protein may be deleted without any adverse effects on cell growth, whereas carboxyl-terminally deleted alleles are unable to support life. Strains with amino-terminally deleted alleles as the only source of TAF61 show no phenotypic abnormalities at low (15°C) or high (37°C) temperature or on medium containing galactose, raffinose, or copper sulfate.

DISCUSSION

We have identified five essential yeast genes and the previously known yeast gene *FUN81* as bona fide yeast TAFs. Two of these (TAF23 and TAF61) have been independently identified in TBP-containing complexes as proteins with apparent molecular masses of 25 and 68 kDa, respectively (9, 11). Previously reported preparations of TBP-associated factors (7, 8) also contain proteins with apparent molecular masses that could be consistent with those of TAF40 and TAF67. Small molecular weight proteins corresponding to TAF17 and TAF19/*FUN81*

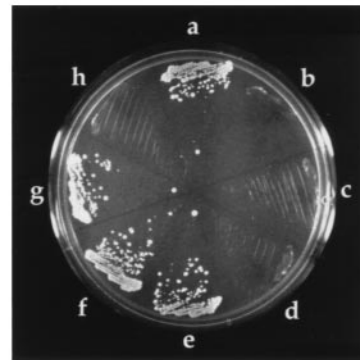
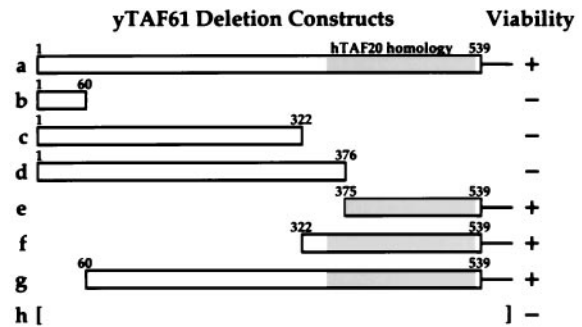


FIG. 5. The carboxyl-terminal domain of yeast TAF61 is necessary and sufficient to support viability. Plasmid shuffling was used to introduce constructs containing the indicated amino acids into a yeast strain with a chromosomal deletion of the *TAF61* gene. Shading indicates the region of the protein with similarity to human TAF₁₁₂₀ and *Drosophila* TAF30 α .

Table 1. TFIID subunit homologies across species

Yeast	Human	<i>Drosophila</i>
TSM1		150
TAF130/145	250	230
None	135	110
TAF90	100	80
TAF67	55	
TAF61 (68)	20/15	30 α (p28/p22)
TAF60	70	60 (62)
TAF40	28	30 β
TAF30 (TFG3/ANC1)	AF-9/ENL	
TAF23 (25)	30	
TAF19 (FUN81)	18	
TAF17	32	40 (42)

Cloning of the indicated TAFs is described in refs. 9, 11, 20, 32, 36, and 38, or reviewed in ref. 2.

were not reported, but they may not have been visible in the gel systems used.

With one exception (see below), every published higher eukaryotic TAF has a unique, essential yeast homologue (Table 1). Human TAF_{II}18 is similar to two yeast genes: *TAF19/FUN81* and *SPT3* (20). However, unlike *TAF19/FUN81*, *SPT3* is not essential for yeast viability (39) and is less similar to the human protein. Therefore, it is clear that yeast TAF19 fulfills a role that is not completely redundant with that of Spt3. The one higher eukaryotic TAF for which there is no yeast homologue is *Drosophila* TAF_{II}110, which has been implicated by *in vitro* experiments in the response to the glutamine-rich activator SP1 (29). The absence of a yeast homologue to *Drosophila* TAF_{II}110 is in accord with the inability of glutamine-rich activators to function in yeast (40) and suggests that perhaps a lack of appropriate coactivator(s) prevents this class of activators from functioning in yeast. We note that the yeast TAF61 sequence contains long polyglutamine stretches reminiscent of those found in dTAF_{II}110; however, these sequences can be completely deleted with no obvious effects on cell growth (Fig. 5).

In fact, the only region of TAF61 necessary for cell growth is the carboxyl-terminal region, which is homologous to the much smaller higher eukaryotic TAFs and to histone H2B. The histone-like structure of some TAFs has been proposed to be functionally important, perhaps in TFIID assembly (36, 41) or in DNA wrapping around TFIID (2). Our observation that this is the essential domain of the yeast TAF61 protein supports the idea of a critical role for this histone-like motif in TAF function.

The TFIID TAF subunits are widely believed to mediate transcriptional activation. The ability of yeast cells to activate transcription after TAF depletion (10, 11) might suggest that other proteins have functional redundancy with TAFs *in vivo*. However, the finding that each higher eukaryotic TFIID subunit has a single homologue in the yeast genome implies that any functional redundancy in response to activators is not due to multiple copies of TAF genes. Because the TAF genes are essential for viability, TAFs must also have other unique critical functions in the cell. These functions remain to be determined.

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